

PROJECT SNOW STORM
LIVE SNOWY OWL SAMPLE SUBMISSION FORM

Band number: _____

Other ID number: _____

(always assign an accession number – either the system your organization uses or following example: 18-cpd-01 (year - your 3 initials - and the sequential number of owls)

Your name: _____

Organization: _____

Your telephone number: _____

Your email address: _____

Captured on: Date: ___/___/___

Time: _____ AM/ PM (circle one)

Capture Location: _____

Released on: Date: ___/___/___

Time: _____ AM/ PM (circle one);

OR

Not yet released (*circle if applicable*); Reason: _____

Transmitter applied: Y N (*circle one*)

Number: _____

Physical Examination:

Fat Index - Check for fat in subcutaneous axillary region, furcular region (V-shaped area on neck between clavicles from which head protrudes), and over abdomen (*circle one of the following*):

- 0 (No visible fat in axillary or abdominal region; furcular region is hollow and completely concave)
- 1 (Trace amount of fat in axillary and abdominal region; the furcular region is concave and the clavicles are visible)
- 2 (Small amount in axillary and abdominal region; furcular region is slightly concave with fat lining clavicles)
- 3 (Moderate amount of visible fat in most regions; furcular region is flat or fat is protruding beyond the clavicles)
- 4 (Obvious fat in axillary and abdominal regions; furcular region full to bulging)
- 5 (Large amount of fat in all regions; abdomen is bulging; fat in furcular region extends past plane of sternum)

Weight(grams): _____

Estimated age: Hatch Year, After Hatch Year (*circle one*); or age: _____ yr;

How determined: _____

Sex: Male Female Unknown (*Circle one*)

How determined: _____

Body Condition Index (BCI): (*circle one*):

- 1 (Emaciated, keel sharp and pointed, little breast muscle)
- 2 (keel easily palpated, reduced and concave breast muscle on either side)
- 3 (keel palpable with light pressure but muscle fills space on either side)
- 4 (keel palpable with firm pressure, muscle is convex on either side of keel)
- 5 (keel not palpable, muscle bulges outward on either side of keel)

BODY SCORE	DESCRIPTION
1	Muscle contour is concave
2	Muscle contour is flat
3	Muscle contour is convex
4	Muscle contour is vesicular
5	Muscle extends beyond keel

Any ectoparasites noted: (circle one)

0= none,

1= 1-2

2= few (3-5)

3= moderate (5-10)

4= heavy (10-15)

5= too numerous to count (> 15)

Body palpation / inspection (Inspect the body - head to toe. Enter NSL (no significant lesions) in blank before each section to indicate that each was examined or describe abnormality or injury):

___ *Head* - Starting with the head - feel all surfaces, noting any abnormalities, abrasions, bumps or irregular areas, look for asymmetry, note featherless areas, swellings, stress bars, etc.

___ *Eyes* - Check the eyes for abnormalities, irregularities of the eyelids, ulcerations or abrasions of the cornea, discharge, asymmetry; record eye color.

___ *Ears* - check each ear for any signs of abnormalities, blood, parasites, debris, etc.

___ *Beak* - check for nasal discharge, abrasions, fractures, misalignment or overgrowth

___ *Oral cavity* - check mouth, under tongue, roof of mouth for abrasions, ulcerations, parasites

___ *Cervical region* - palpate neck region for any abnormalities, swellings, etc.

___ *Thorax* - palpate chest and back for any abnormalities, bumps, etc.

___ *Wings* - extend wings to check for full extension, palpate for abnormalities, bumps, old fractures

___ *Abdomen* - palpate abdomen for any abnormalities, lumps, etc.

___ *Cloaca* - check for signs of diarrhea - matted dirty feathers at anal opening

____ *Uropygial gland* - inspect for any abnormalities - swelling, etc.

____ *Legs/feet* - palpate for abnormalities, signs of old fractures, check feet for missing toes, bumble foot

____ *Feathers* – note any feather damage (tipped or broken feathers, stress bars), check for missing feathers, or new/pin feathers (molt stage), and note any feather soiling

Blood collection:

Instructions:

1. Use a 6 or 3 ml syringe and a 22 ga needle to collect a blood sample (ideally 3-5 ml total) from the right jugular vein (ideally) or the left or right ulnar (wing) vein or the left or right medial metatarsal (inner leg) vein (jugular vein will yield a much larger sample to allow all needed testing; will likely need smaller needle and syringe for other locations).
2. Place blood sample immediately into at least 2 green top (sodium or lithium heparin without gel and without rubber stoppers) tube(s) and gently rotate several times.
3. Make 2-4 blood smear slides immediately after placing into tube (within 30 minutes) and allow to air dry.
4. One of the green top tubes (1.5-2.0 ml) should remain unspun as is and placed into refrigerator until shipment. DO NOT FREEZE.
 - a. If there are serious shipping delays that will impact analysis quality (longer than 3 days), and a local source competent in doing CBC samples on birds is available (e.g., IDEXX, Antech, other local laboratories), samples can be analyzed locally and Project SNOWstorm will reimburse reasonable analysis costs. Please include results with samples or send to Dr. Ellen Bronson via email (contacts below).
5. Remainder of blood tubes should be centrifuged and the plasma decanted into tubes such as cryovials without anticoagulant. (need ideally at least 2.0 ml but will work with any amount you can send). Place sample on ice or freeze at first opportunity.
6. Deliver slides, refrigerated whole blood and frozen plasma to Maryland Zoo lab via Federal Express overnight on ice packs; only send Monday through Thursday to the following address:
Veterinary Hospital Lab
Maryland Zoo in Baltimore
1876 Mansion House Drive
Baltimore, MD 21217

Please mark package as “Perishable”

Please contact Dr. Ellen Bronson (see contacts below) before shipping to confirm timing.

Blood collection: Date: ___/___/___ Time: _____ AM/ PM (circle one)

Vessel used: jugular ulnar leg (circle one)

Spun and separated: Date: ___/___/___ Time: _____ AM/ PM (circle one)

Frozen (if applicable): Date: ___/___/___ Time: _____ AM/ PM (circle one)

Shipped overnight on: Date: ___/___/___ Time: _____ AM/ PM (circle one)

Other samples:

Fecal sample (if bird defecates brown or green during restraint, place in clean vial, keep cool not frozen, and send with blood sample)

Avian Influenza SWABS:

Use sterile synthetic or semi-synthetic swabs (e.g., polyester, rayon, nylon) with a plastic handle (flocked or spun head). Avoid cotton or calcium alginate swabs or swabs with wooden handles.

Swab the oropharynx (*swirl swab in back of oral cavity*) and cloaca (*place swab gently into cloaca and swirl*) with same swab; place in 1 vial in -80 C freezer; if possible place in preservative/ media such as BHI. (*your state department of agriculture, state wildlife agency veterinarian, or university may be able to provide you with swabs and media vials*). Send with the blood sample cooled; if more than 2 days, freeze and send on ice with other frozen samples.

Biosecurity / Clean up:

* Place needles, blood tube waste, and glass slide waste in sharps disposal; place syringes in waste bag

* Wipe all surfaces and equipment and tools with disinfectant

For further information please contact any one of the following three veterinarians who will be happy to answer questions and provide direction to assist with obtaining the best quality samples.

1st) Dr. Ellen Bronson
Senior Veterinarian
Maryland Zoo Hospital
1876 Mansion House Drive
Baltimore, Maryland 21217
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Cell: 443-823-3691
Email: ellen.bronson@marylandzoo.org

2nd) Dr. Erica Miller
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3rd) Dr. Cindy Driscoll
Maryland State Wildlife Veterinarian
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Oxford, Maryland 21654
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